

***In vivo* volumetric imaging of subcutaneous microvasculature by photoacoustic microscopy**

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Abstract: Photoacoustic microscopy was developed to achieve volumetric imaging of the anatomy and functions of the subcutaneous microvasculature in both small animals and humans *in vivo* with high spatial resolution and high signal-to-background ratio. By following the skin contour in raster scanning, the ultrasonic transducer maintains focusing in the region of interest. Furthermore, off-focus lateral resolution is improved by using a synthetic-aperture focusing technique based on the virtual point detector concept. Structural images are acquired in both rats and humans, whereas functional images representing hemoglobin oxygen saturation are acquired in rats. After multiscale vesselness filtering, arterioles and venules in the image are separated based on the imaged oxygen saturation levels. Detailed structural information, such as vessel depth and spatial orientation, are revealed by volume rendering.

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References and links

1. R. L. Barnhill, K. Fandrey, M. A. Levy, M. C. Mihm, and B. Hyman, "Angiogenesis and tumor progression of melanoma: quantification of vascularity in melanocytic nevi and cutaneous malignant melanoma," *Lab. Invest.* **67**, 331-337 (1992).
2. H. Nakajima, T. Minabe, and N. Imanishi, "Three-dimensional analysis and classification of arteries in the skin and subcutaneous adipofascial tissue by computer graphics imaging," *Plast. Reconstr. Surg.* **102**, 748-760 (1998).
3. X. Wang, Y. Pang, G. Ku, X. Xie, G. Stoica, and L. V. Wang, "Noninvasive laser-induced photoacoustic tomography for structural and functional *in vivo* imaging of the brain," *Nat. Biotechnol.* **25**, 114-116 (2000).
4. R. R. Edelman, H. P. Mattle, D. J. Atkinson, and H. M. Hoogewoud, "MR angiography," *Am. J. Roentgenol.* **154**, 937-946 (1990).
5. Y. Zhao, Z. Chen, C. Saxer, S. Xiang, J. F. de Boer, and J. S. Nelson, "Phase-resolved optical coherence tomography and optical Doppler tomography for imaging blood flow in human skin with fast scanning speed and high velocity sensitivity," *Opt. Lett.* **25**, 114-116 (2000).
6. F. A. Duck, *Physical Properties of Tissue* (Academic Press, San Diego, CA, 1990).
7. C. G. A. Hoelen, F. F. M. de Mul, R. Pongers, and A. Dekker, "Three-dimensional photoacoustic imaging of blood vessels in tissue," *Opt. Lett.* **23**, 648-650 (1998).
8. M. C. Pilatou, N. J. Voogd, F. F. M. de Mul, and W. Steenbergen, "Analysis of three-dimensional photoacoustic imaging of a vasculature tree *in vitro*," *Rev. Sci. Instrum.* **74**, 4495-4499 (2003).
9. R. G. Kolkman, E. Hondebrink, W. Steenbergen, and F. F. M. de Mul, "*In vivo* photoacoustic imaging of blood vessels using an extreme-narrow aperture sensor," *IEEE J. Sel. Top. Quantum Electron.* **9**, 343-346 (2003).
10. X. Wang, Y. Pang, G. Ku, G. Stoica, and L. V. Wang, "Three-dimensional laser-induced photoacoustic tomography of mouse brain with the skin and skull intact," *Opt. Lett.* **28**, 1739-1741 (2003).
11. M. Xu, and L. V. Wang, "Photoacoustic imaging in biomedicine," *Rev. Sci. Instrum.* **77**, 041101 (2006).

12. A. A. Oraevsky, and A. A. Karabutov, "Optoacoustic Tomography," in *Biomedical Photonics Handbook*, T. Vo-Dinh ed. (CRC Press, Boca Raton, FL, 2003).
 13. M. Xu, and L. V. Wang, "Analytic explanation of spatial resolution related to bandwidth and detector aperture size in thermoacoustic and photoacoustic reconstruction," *Phys. Rev. E* **67**, 1-15 (2003).
 14. K. Maslov, G. Stoica, and L. V. Wang, "*In vivo* dark-field reflection-mode photoacoustic microscopy," *Opt. Lett.* **30**, 625-627 (2005).
 15. H. F. Zhang, K. Maslov, G. Stoica, and L. V. Wang, "Functional photoacoustic microscopy for high-resolution and noninvasive *in vivo* imaging," *Nature Biotechnol.* **24**, 848-851 (2006).
 16. National Institutes of Health, "Guide for the Care and Use of Laboratory Animals," (U.S. Government Printing Office, Washington DC, 1985), NIH Pub. 86-23.
 17. M.-L. Li, H. F. Zhang, K. Maslov, G. Stoica, and L. V. Wang, "Improved in-vivo photoacoustic microscopy based on a virtual detector concept," *Opt. Lett.* **31**, 474-476 (2006).
 18. D. Jensen, *The principles of physiology* (Appleton-Century-Crofts, New York 1976), pp 746.
 19. K. Maslov, M. Sivaramakrishnan, H. F. Zhang, G. Stoica, and L. V. Wang, "Technical considerations in quantitative blood oxygenation measurement using photoacoustic microscopy in small animal *in vivo*," in *Photons Plus Ultrasound: Imaging and Sensing 2006*, A. A. Oraevsky and L. V. Wang, eds., Proc. SPIE **6086**, 215-225 (2006).
 20. A. F. Frangi, W. J. Niessen, K. L. Vincken, M. A. Viergever, "Multiscale vessel enhancement filtering," in *Proceedings of Medical Image Computing & Computer Assisted Intervention (MICCAI)*, W. Wells, A. Colchester, and S. Delp, eds., Vol. 1496 of Lecture Notes in Computer Science, (Springer-Verlag, Berlin 1998), pp.130-137.
 21. American national standard for the safe use of lasers Z136.1, (American National Standards Institute, New York, 2000).
 22. P. Carmeliet and R. K. Jain, "Angiogenesis in cancer and other disease," *Nature* **407**, 249-257 (2000).
 23. C. M. van Vommel, L. J. Spreeuwiers, M. A. Viergever, and W. J. Niessen, "Level-set-based artery-vein separation in blood pool agent CE-MR angiograms," *IEEE Trans. Med. Imaging.* **22**, 1224-1234 (2003).
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1. Introduction

High-resolution imaging of subcutaneous blood microvessels in three dimensions can be of great benefit to dermatology, cancer research [1], and plastic surgery [2]. However, accurate noninvasive imaging of the anatomy and functions of the subcutaneous microvasculature remains a challenge. First, the imaging modality employed must have high sensitivity so that small-diameter (tens of μm) blood vessels can be detected. Second, the imaging modality employed must have high spatial resolution so that the vessel network can be resolved. Third, it must have enough penetration depth so that the major vasculature in the dermal layer (1-3 mm below the skin surface in humans and small animals) can be measured. Fourth, it must be able to acquire volumetric data with good spatial resolution in both the lateral and axial directions. Fifth, it must image appropriate intrinsic contrast so that physiological functions can be measured.

In blood vessel imaging, the intrinsic contrast comes from either the strong optical absorption of hemoglobin [3] or the velocity of the blood flow [4,5]. Because the relative optical absorption contrast between blood and dermal tissue can be up to 100, depending on the optical wavelength used [6], an optical absorption-based high-resolution imaging modality is well suited for microvascular imaging. Moreover, since optical absorption is associated with both forms of hemoglobin in blood, optical absorption-based imaging can quantify functional parameters such as the total hemoglobin concentration and hemoglobin oxygen concentration (SO_2).

The ratio of the maximum imaging depth to the depth (axial) resolution, which gives the number of effective pixels, is used to evaluate the spatial resolution and image quality of an imaging modality. A depth-to-resolution ratio greater than 100 has been achieved by several optical modalities, such as confocal microscopy, two-photon microscopy, and optical coherence tomography (OCT). However, all are not directly dependent on optical absorption contrast, and none can reach a penetration depth of more than ~ 1 mm (related to one transport mean free path) in biological tissue owing to strong optical scattering. Hence, existing high-resolution optical imaging modalities are not ideal for imaging subcutaneous microvasculatures.

